Author's response to reviews

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Evaluation of Phytochemical content, antimicrobial, cytotoxic and antitumor activities of extract from *Rumex hastatus* D.Don roots

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Abstract

Background:
Being a part of Chinese as well as ayurdic herbal system, roots of *Rumex hastatus* D.Don (RH) is highly medicinal, used to regulated blood pressure. It is also reported that the plant is diuretic, laxative, tonic, used against microbial skin diseases, bilious complaints and jaundice. The present study is conducted to evaluate phytochemical, antimicrobial, antitumor and cytotoxic activities of extract obtained from *R. hastatus* roots.

Methods:
RH roots were powdered and extracted with methanol to get crude extract. Crude extract was further fractioned on the basis of increasing polarity, with n-hexane (HRR), chloroform (CRR), ethyl acetate (ERR), n-butanol (BRR) and residual aqueous fraction (ARR). Methanol extract and its derived fractions were subjected to phytochemical screening and assayed for antibacterial activities via agar well diffusion method. Antifungal activities were checked through agar tube dilution method whereas potato disc assay was employed for the determination of antitumor activity. On the other hand cytotoxic activities were conducted using brine shrimps procedures.

Results:
The results obtained from phytochemical analysis indicate the presence of **alkaloids, anthraquinones, flavonoids and saponins** in all the fractions. Most of the plant fractions showed substantial antimicrobial activities, which is in accordance with the spacious use of tested plant samples in primary healthcare center. Fractions of *R. hastatus* roots for cytotoxicity were tested as an effective cytotoxic was found as BRR>MRR>CRR>ARR>ERR> HRR. Ranking order of fractions of *R. hastatus* roots for effective antitumor screening was found as MRR >BRR >ARR> CRR >ERR >HRR.
Conclusion:

These results showed that *R. hastatus* appeared as an important source for the discovery of new antimicrobial drugs and antitumor agents; verify its traditional uses and its exploitation as therapeutic agent.

**Keywords:** *Rumex hastatus*, roots, phytochemicals, antimicrobial, cytotoxic, antitumor
Several countries use plants as a best source of medicine to treat infections and other disorders and nowadays most potent and powerful drugs are derived from plants [1]. Natural products derived from plant extracts/fractions are novel therapeutic agents for various infectious as well as degenerative diseases. In herbal medicines various parts of the plant (root, stem, flower, fruit, twigs exudates and modified plant organs) are used having diverse therapeutic properties. To utilize these plants, collected on minute scale by local communities and folk healers, while to trade for herbal industries numerous other plants are collected in large amount as a raw material [2]. People of developed and developing countries have turned back their attention towards botanicals for a substitutive health care services as they are an accessible and economical source in comparison to synthetic medicines. In recent years for the management and protection against pathogens, a large number of plants have been examined for their antimicrobial characteristics as an integrative system of medicine. For the treatment of human pathogenic diseases, use of plant extracts or fractions having antimicrobial properties can be of great importance. Natural biological active compounds in plants have a significant role in plants defense mechanism and also important for their unambiguous physiological action in human body. Because of the therapeutic property, secondary metabolites (flavonoids, alkaloids, tannins, saponins, and terpenoids) are becoming a part of the integrative health care system as supportive and alternative medicines [3]. For the cure of infectious diseases the unsystematic use of commercial antimicrobial medicines has led to development of numerous drug resistances in human pathogenic microorganisms. Besides, a number of side effects like hypersensitivity, allergic reactions and immune suppression are rarely associated with the antibiotics. Further, food preservation requires evaluation of natural resources such as herbal fractions and isolates with
antimicrobial properties as the long historic use of herbs has proved their safety and efficacy in
various traditional medicine systems. Recent trends for the use of natural remedies as
antimicrobial have increased their use in food, cosmetic and pharmaceutical products which have
been screened in vitro and indicated antimicrobial and other diverse properties [4]. Novel
compounds have a vast therapeutic ability, lessen various adverse effects, and are often coupled
with synthetic antimicrobials. One of the valuable means used to screen bioactive compounds
from plant extracts is brine shrimp lethality assay [5]. It is considered as a convenient probe for
primary evaluation of toxicity [6]. Medicinal plants are good source to obtain a wide range of
drugs in view of the fact that, a single plant can be used to treat more than one ailment. *Rumex*
hastatus* D. Don, belongs to the Polygonaceae family. The plant is suffrutescent richly branching
shrub. It grows up to 90-120 cm tall and leaves with petioles of the same length as the blade;
blade hastate, panicles terminal with erect-divergent, mostly simple branches, nut up to 2 mm
long, brown, and long spindle-shaped roots. It is distributed in northern Pakistan, north east
Afghanistan and south west of China, growing between 700-2500 m, sometimes grows as pure
population [7]. The present study is therefore arranged to screen and evaluate antibacterial,
antifungal, antitumor and cytotoxic activities as well as phytochemical analysis of crude
methanol extract.

**Methods**

**Extract preparation**

*R. hastatus* District roots were collected from Havelian, Abbottabad, Pakistan. The plant
specimen are documented by their native names and then authenticated by Prof. Dr. Mir Ajab
Khan, Department of Plant Sciences, Quaid-i-Azam University, Islamabad. Voucher specimens
with Accession No. 27813 (R. hastatus) were placed at the Herbarium of Pakistan. Shade dried roots were powdered in a Willy Mill to 60-mesh size and used for solvent extraction. Five kilogram of each powdered sample was extracted twice with 10 L of 95% methanol at 25 °C for 48 h. For filtration Whatman No. 1 filter paper was used and then the filtrate was concentrated on rotary evaporator (Panchun Scientific Co., Kaohsiung, Taiwan) under reduced pressure at 40°C. In order to resolve the compounds with escalating polarity, a part of the extract was suspended in distilled water and partitioned with n-hexane (HRR), chloroform (CRR), ethyl acetate (ERR), n-butanol (BRR) and residual aqueous fraction (ARR).

**Determination of the phytochemical constituents**

The extract of R. hastatus roots and its derived fractions were evaluated for the occurrence of alkaloids, saponins and terpenoids [8], whereas anthraquinone, phlobatannins and cardiac glycosides [9], as well as flavonoids and tannins [10], using standard phytochemical methods.

**Antimicrobial activities**

All the microbes obtained from Department of Microbiology QAU, Islamabad, Pakistan while antibiotics were obtained from GlaxoSmithKline however the thymol and camptothecin are purchased from Sigma.

**Antibacterial activity**

For antibacterial activity the strains used were Bacillus subtilis (ATCC 6633), Enterobacter aerogenes (ATCC 13048), Escherichia coli (ATCC 15224), Klebsiella pneumoniae (MTCC 618), Micrococcus luteus (ATCC 10240), Pseudomonas aeruginosa (ATCC 27853), Salmonella typhi (ATCC 0650), and Staphylococcus aureus (ATCC6538). Antibacterial activity of plant
extract and its derived fractions was investigated by agar well diffusion method [11] using nutrient agar medium. Extract and fractions of different concentration were prepared in DMSO. Erythromycin and nalidixic acid were used as positive controls whereas DMSO was used as a negative control. The lowest concentration inhibiting growth was taken as the minimum inhibitory concentration (MIC).

Antifungal activity

The agar tube dilution method was used for antifungal activity of plant extracts was determined according to the protocol reported by Duraipandiyan and Ignacimuthu [12] using Aspergillus flavus (0064), Aspergillus fumigatus (66), Aspergillus niger (0198) and Fusarium solani (0300). Tubes were prepared in triplicate for each fungus species. Percentage inhibition of fungal growth for each concentration of fractions was determined by the following formula;

\[
\text{Percentage inhibition of fungal growth} = \left( 100 - \frac{\text{linear growth in test (mm)}}{\text{linear growth in control (mm)}} \right) \times 100
\]

Cytotoxic brine shrimp assay

*R. hastatus* and its various fractions were analyzed by using brine shrimps hatched in saline [13]. 10, 100, 1000 ppm was incubated with brine shrimps. After 24 h of incubation survivors were counted with help of 3x magnifying glass and calculation was done using Abbot’s formula;

\[
\% \text{Death} = \left( \frac{\text{Sample} - \text{control}}{\text{control}} \right) \times 100
\]

Antitumor potato disc assay

*R. hastatus* and its derived fractions were analyzed for their antitumor activity by antitumor potato disc assay reported by Ferrigini *et al.*, [14]. Bacterial culture of *Agrobacterium*
*tumefaciens* (At 10) was used. Number of tumors per disc was counted and percentage inhibition for each concentration was determined as follows:

\[
\text{Percentage inhibition} = 100 - \left( \frac{\text{average number of tumors of sample}}{\text{Average number of tumors of negative control}} \right) \times 100
\]

20% tumor inhibition was considered significant.

**Statistical analysis**

Data of *in vitro* assays recorded were analyzed with help computerized Graph prism pad software to determined standard error, IC\(_{50}\) and LD\(_{50}\).

**Results and discussion**

Plants and plant derived fractions are used for medication since prehistory. As reported by World Health Organization (WHO) about 80% of the world’s population depends mainly on conventional medicines that involve the use of plant extracts [15-16]. The folkloric use of *R. hastatus* for treating infections may be justified by the charisma of anthraquinones as they possess antimicrobial [17] and antiparasitic properties [18]. Coumarins are known as antioxidant, anti-inflammatory, antiviral, anticarcinogenic and hepatoprotective agent like hydroxycoumarins have ability to chelate metal ions and scavenge free radicals [19]. Present study supports the examined plant parts as large sources of bioactive chemicals specifically with reference to alkaloids, anthraquinones, coumarins, flavonoids, tannins and saponins that ought to be isolated and monitored for biological activities as reported in traditional and therapeutic utilization [19].

In the present study *R. hastatus* roots showed alkaloids, anthraquinones, flavonoids and saponins were present in MRR and its various fractions such as HRR, ERR, CRR, BRR and ARR (Table 1). Presence of tannins was not detected in HRR and ARR while terpenoids were remained
absent in HRR and ERR. Cardiac glycosides were present in MRR, ERR and BRR while coumarins were not reported in ERR and ARR. Analysis of phlobatannins revealed their presence only in MRR and BRR.

In the present study different bioassays were used to characterize the plant fractions. The MIC measurement is a quantitative method, corresponds to the lowest applied amount of test material able to inhibit any visible microbial growth. According to Lambert and Pearson [20], MIC is a standard measure documented for the susceptibility of organisms to inhibitors in biological assays. The antibacterial activity was tested against both gram-negative bacteria as well as gram-positive bacteria. Table 2 describes the antibacterial activity as MIC value of various fractions of plant samples against tested bacteria. The results obtained from the present study for all tested bacteria ranged from 0.1 to 10 mg/ml. Gram-positive bacteria such as *Staphylococcus aureus* was inhibited by MIC value (1 mg/ml) of MRR; 2.5 mg/ml of HRR, BRR; 5 mg/ml of CRR while rest of the fractions didn’t inhibit the growth of *Staphylococcus aureus*. On the other hand, *Bacillus subtilis* was inhibited by ERR (0.1 mg/ml); MRR (1 mg/ml); BRR (0.5 mg/ml); ARR (2.5 mg/ml); CRR (5 mg/ml) while remaining fractions did not inhibit the growth of the respective bacteria (Table 2). In case of gram negative bacteria *Klebsiella pneumoniae*’s growth was inhibited by MRR (5 mg/ml); HRR, ERR (10 mg/ml) while remaining fractions did not inhibit the growth of the respective bacteria. *Pseudomonas aeruginosa* inhibited the growth at 0.1 mg/ml. Growth of *Salmonella typhi* was inhibited by HRR (0.5 mg/ml); BRR (0.25 mg/ml); while rest of the fractions inhibited the growth at 0.1 mg/ml. However, BRR inhibited the growth of *Enterobacter aerogenes* with MIC (1 mg/ml); CRR, MRR (2.5 mg/ml); ARR (5 mg/ml); while remaining fractions did not show inhibition of the concerned bacteria (Table 2). MIC against *Micrococcus luteus* and *Escherichia coli* had no value, showing no antibacterial activity.
of any of the fractions. The activity against both type of bacteria (gram-positive and gram-negative bacteria) indicate that fractions contain broad spectrum of antibiotic compounds or metabolic toxins. These fractions may possibly be functional for the strengthening of new antimicrobial drugs. Numbers of authors have focused on antibacterial and antifungal potency of flavonoids [21-22]. Antimicrobial activity may be attributed to plant bioactive compounds to make complex with bacterial cell wall [17] and thus inhibiting the microbial growth.

Our results suggest that antimicrobial activity of plant fractions doesn’t depends on only phenolics but other secondary metabolites are also involved. While, R. hastatus roots showed that inhibition against A. niger ranged from 20.4±2.85% to 68.3±1.48%, A. flavus at the range from 23.6±1.32 % to 69.4±3.18 %, A. fumigatus at a range from 15.5±2.34% to 51.3±1.67% and F. solani at a range from 14.1±1.51% to 34.9±2.28% (Table3).

For the investigation of bioactive compounds, biological assays especially brine shrimp lethality assay (BSLA) is considered as necessary and suitable tools. According to Meyer et al. [24] who categorized crude extracts and pure compounds into toxic (LC50 value < 1000 ppm) and non-toxic (LC50 value > 1000 ppm), all tested showed good brine shrimp larvicidal activity excluding HRR. Moreover, Peteros and Uy [23] signified the presence of effective cytotoxic substances of plant extracts having LC50 values < 100 ppm to brine shrimp lethality.

Our findings were in strong accordance to that of Peteros and Uy [23] and suggest isolating new active compounds. R. hastatus roots for cytotoxicity were tested and their ranking order for effective cytotoxic was found as BRR > MRR > CRR > ARR > ERR > HRR (Table4). About all of the tested samples showed their toxic effect at 1000 ppm apart HRR. The larvicidal activity of R. hastatus were in consistent with the Hussain et al. [25] who reported the cytotoxic activity of methanol extracts of Rumex species and found that R. hastatus showed significant activity.
against brine shrimp larvae. It is considered that presence of a wide range of biologically active compounds with different structures and their synergistic effects may add to the overall activity of particular fraction.

Antitumor potato disc is one of the best bioassay used for the detection of biologically active components in the botanical extracts [26]. Tumor inducing plasmid multiplies the plant’s cells excluding apoptosis phase and ultimately tumor is formed that has similarity with human and animal cancers, histology and nucleic acid [27-28]. Several scientists have been used these methods over the past 15 years, and they appear to be adaptable to the purpose of standardization or quality control of bioactive compounds in such heterogeneous botanicals [29]. Table 5 describes that the ranking order for effective antitumor screening of fractions of R. hastatus roots was found as MRR > BRR > ARR > CRR > ERR > HRR. Different strains can be used to induce tumor on potato discs but strain Atl0 was found more prominent for producing tumor [30-31]. Our results are in accord to other studies [32] who reported that tumor inhibition rates on potato discs are dependent on the concentration of the samples. Our findings confirm the previous reports of Islam et al. [31] and Ashraf et al. [33] who reported that the antitumor activity perhaps endorsed with the nature of biological active compounds and their strong solubility with appropriate solvent and also justifying the statement of Fatima et al. [32] that tumor induction was variable in case of different solvent extracts.

**Conclusion**

The result of this study indicates the potential of R. hastatus roots as a source of therapeutic agent against microbial infection and tumor diseases.

**Competing interests**

The authors declare that they have no competing interests.
Authors’ contributions
SS made significant contribution to acquisition of data, analysis, drafting of the manuscript.
MRK and RAK (ORCID ID: 0000-0003-0453-2090) have made substantial contribution to conception and design, interpretation of data, drafting and revising the manuscript for intellectual content. TAH participated in the analysis design and drafting of the manuscript. All authors read and approved the final manuscript.

Acknowledgment
We acknowledge HEC Pakistan for fund provision in the compilation of the research work.

References


<table>
<thead>
<tr>
<th>Phytochemical constituents</th>
<th>MRR</th>
<th>HRR</th>
<th>ERR</th>
<th>CRR</th>
<th>BRR</th>
<th>ARR</th>
</tr>
</thead>
<tbody>
<tr>
<td>Alkaloids</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Anthraquinones</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Cardiac glycosides</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>Coumarins</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>Flavonoids</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Phlobatannins</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>Saponins</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>Tannins</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>Terpenoids</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
</tbody>
</table>

+, present; -, absent

MRR; *R. hastatus* methanol extract, HRR; *R. hastatus* n-hexane fraction, CRR; *R. hastatus* chloroform fraction, ERR; *R. hastatus* ethyl acetate fraction, BRR; *R. hastatus* n-butanol fraction, ARR; *R. hastatus* residual aqueous fraction.
Table 2 The Minimum inhibitory concentration (mg/ml) of *R. hastatus*

<table>
<thead>
<tr>
<th>Bacteria</th>
<th>MRR</th>
<th>HRR</th>
<th>ERR</th>
<th>CRR</th>
<th>BRR</th>
<th>ARR</th>
<th>Erythromycin</th>
<th>Nalidixic acid</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>B. subtilis</em></td>
<td>1.0±0.01</td>
<td>-</td>
<td>0.1±0.01</td>
<td>5.0±0.5</td>
<td>0.5±0.01</td>
<td>2.5±0.07</td>
<td>0.006±0.0001</td>
<td>0.001±0.0007</td>
</tr>
<tr>
<td><em>E. aerogenes</em></td>
<td>2.5±0.02</td>
<td>-</td>
<td>-</td>
<td>2.5±0.3</td>
<td>1.0±0.01</td>
<td>5.0±0.3</td>
<td>0.007±0.0002</td>
<td>0.002±0.001</td>
</tr>
<tr>
<td><em>E. coli</em></td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>0.009±0.0002</td>
<td>0.07±0.009</td>
</tr>
<tr>
<td><em>K. pneumoniae</em></td>
<td>5.0±0.3</td>
<td>10.0±0.2</td>
<td>10.0±0.9</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>0.004±0.0002</td>
<td>0.001±0.0004</td>
</tr>
<tr>
<td><em>M. luteus</em></td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>0.006±0.0001</td>
<td>0.001±0.0003</td>
</tr>
<tr>
<td><em>S. typhi</em></td>
<td>0.1±0.001</td>
<td>0.5±0.01</td>
<td>0.1±0.01</td>
<td>0.1±0.01</td>
<td>0.25±0.1</td>
<td>0.1±0.01</td>
<td>0.001±0.0002</td>
<td>0.007±0.0008</td>
</tr>
<tr>
<td><em>P. aeruginosa</em></td>
<td>0.1±0.002</td>
<td>0.1±0.01</td>
<td>0.1±0.07</td>
<td>0.1±0.02</td>
<td>0.1±0.01</td>
<td>0.1±0.01</td>
<td>0.003±0.0001</td>
<td>0.001±0.0001</td>
</tr>
<tr>
<td><em>S. aureus</em></td>
<td>1.0±0.07</td>
<td>2.5±0.1</td>
<td>-</td>
<td>5.0±0.6</td>
<td>2.5±0.01</td>
<td>-</td>
<td>0.007±0.0002</td>
<td>0.001±0.0009</td>
</tr>
</tbody>
</table>

Mean±SE (n=3)

- = not active against tested microorganism.
Table 3 Antifungal activity of the crude extract and various fractions of *R. hastatus* (Percent inhibition)

<table>
<thead>
<tr>
<th></th>
<th>MRR</th>
<th>HRR</th>
<th>ERR</th>
<th>CRR</th>
<th>BRR</th>
<th>ARR</th>
<th>Terbinafin</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>A. niger</em></td>
<td>39.1±1.39</td>
<td>40.2±2.83</td>
<td>48.9±1.66</td>
<td>68.3±1.48</td>
<td>28.4±2.51</td>
<td>20.4±2.85</td>
<td>84.5±2.13</td>
</tr>
<tr>
<td><em>F. solani</em></td>
<td>25.4±2.53</td>
<td>15.8±1.39</td>
<td>14.1±1.51</td>
<td>31.7±2.49</td>
<td>31.7±2.49</td>
<td>31.7±2.49</td>
<td>81.78±2.12</td>
</tr>
<tr>
<td><em>A. flavus</em></td>
<td>69.4±3.18</td>
<td>36.4±1.89</td>
<td>28.0±1.34</td>
<td>36.0±1.68</td>
<td>56.2±2.77</td>
<td>23.6±1.32</td>
<td>87.6±2.70</td>
</tr>
<tr>
<td><em>A. fumigatus</em></td>
<td>22.6±2.32</td>
<td>27.0±2.56</td>
<td>43.2±1.80</td>
<td>51.3±1.67</td>
<td>15.5±2.34</td>
<td>40.5±1.07</td>
<td>88.4±3.16</td>
</tr>
</tbody>
</table>

Mean±SE (n=3)
Table 4 Illustration of % age mortality of brine shrimps at different concentrations of extract and fractions and respective LD50 values

<table>
<thead>
<tr>
<th>Extracts</th>
<th>10 ppm</th>
<th>100 ppm</th>
<th>1000 ppm</th>
<th>LD50 (ppm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MRR</td>
<td>1.76±0.05</td>
<td>2.28±0.17</td>
<td>3.65±0.15</td>
<td>60.0±3.5</td>
</tr>
<tr>
<td>HRR</td>
<td>1.05±0.31</td>
<td>2.11±0.02</td>
<td>2.18±0.09</td>
<td>&gt;1000</td>
</tr>
<tr>
<td>ERR</td>
<td>1.66±0.05</td>
<td>1.50±0.03</td>
<td>1.79±0.06</td>
<td>705.5±5.2</td>
</tr>
<tr>
<td>CRR</td>
<td>2.65±0.22</td>
<td>2.13±0.05</td>
<td>1.52±0.12</td>
<td>65.7±4.1</td>
</tr>
<tr>
<td>BRR</td>
<td>2.53±0.31</td>
<td>2.81±0.03</td>
<td>3.05±0.09</td>
<td>15.4±1.5</td>
</tr>
<tr>
<td>ARR</td>
<td>3.15±0.08</td>
<td>3.16±0.06</td>
<td>1.72±0.23</td>
<td>100±2.7</td>
</tr>
<tr>
<td>Thymol</td>
<td>1.62±0.04</td>
<td>2.89±0.07</td>
<td>2.75±0.13</td>
<td>&lt;10±0.07</td>
</tr>
</tbody>
</table>

Mean±SE (n=3)
Table 5  % tumors inhibition of various fractions of *R. hastatus* against potato disc tumor

<table>
<thead>
<tr>
<th>Extracts</th>
<th>10 ppm</th>
<th>100 ppm</th>
<th>1000 ppm</th>
<th>IC50 (ppm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MRR</td>
<td>26.24±1.20</td>
<td>69.33±1.56</td>
<td>87.00±0.85</td>
<td>60</td>
</tr>
<tr>
<td>HRR</td>
<td>19.43±0.75</td>
<td>26.00±0.52</td>
<td>37.47±1.08</td>
<td>&gt;1000</td>
</tr>
<tr>
<td>ERR</td>
<td>35.43±0.38</td>
<td>47.62±0.94</td>
<td>58.17±1.20</td>
<td>340</td>
</tr>
<tr>
<td>CRR</td>
<td>32.47±1.32</td>
<td>56.80±2.17</td>
<td>75.82±2.63</td>
<td>80</td>
</tr>
<tr>
<td>BRR</td>
<td>19.86±0.44</td>
<td>64.36±0.77</td>
<td>84.33±1.43</td>
<td>70</td>
</tr>
<tr>
<td>ARR</td>
<td>24.65±0.63</td>
<td>63.33±2.43</td>
<td>78.33±2.61</td>
<td>70</td>
</tr>
<tr>
<td><strong>Camptothecin</strong></td>
<td><strong>49.2±1.2</strong></td>
<td><strong>69.5±2.3</strong></td>
<td><strong>88.9±3.2</strong></td>
<td><strong>11</strong></td>
</tr>
</tbody>
</table>

Mean±SE (n=3)