Author's response to reviews

Title: In-vitro and in-vivo validation of ethnopharmacological uses of methanol extract of Isodon rugosus Wall. ex Benth. (Lamiaceae)

Authors:

Khalid Hussain Janbaz (khjanbaz@hotmail.com)
Javeria Arif (javeria.arif46@yahoo.com)
Fatima Saqib (fatimasaqib786@yahoo.com)
Imran Imran (imran.ch@bzu.edu.pk)
Muhammad Ashraf (muhammadashraf291@yahoo.com)
Muhammad Zia-Ul-Haq (ahirzia@gmail.com)
Hawa Z.E. Jaafar (hawazej@gmail.com)
Vincenzo De Feo (defeo@unisa.it)

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Author's response to reviews: see over
Dear Editor

BMC Complementary and Alternative Medicine

Subject: Submission of revised MS. 8769346731097018

Dear Sir,

I am highly thankful for referee comments for improving manuscript No. MS: 8769346731097018 entitled, “In-vitro and in-vivo validation of ethnopharmacological uses of Isodon rugosus”. We have amended manuscript as per referee suggestions and we hope now it is in better position. Referee comments really helped in improving the manuscript. Following is specific response to referee comments

Referee 1

Q. The title is must include the solvent used, type of preparation and part of the plant used.

Ans. As per recommendation we modified the title of manuscript.

Q. The doses/ concentrations of the extract used should be indicated in abstract.

Ans. We added the doses of extract in abstract at appropriate position.

Q. The vehicle used for reconstitution of extract is not clearly mentioned throughout.

Ans. In modification, it is now mentioned that “The stock solutions of Isodon rugosus were prepared in distilled water and the dilutions were made fresh in normal saline on the day of experiment.”

Q. The plant collected should have specimen number for references.

Ans. We have added it now.

Q. For extraction, what proportion of solvent and plant material was macerated?
Ans. Now added in the manuscript; “Approximately 500 g powdered plant material was soaked in 80% aqueous-methanol by cold maceration at room temperature for 7 days with occasional shaking”

Q. Authors listed Wistar rats, but no result indicating their use

Ans. It was mistake and now eliminated from the text.

Q. Why authors used isotonic contractions for rabbit jejunum preparations and isometric contractions for rabbit tracheal and aortal preparations?

Ans. These are the standard method established by ADI instruments for the determination of contraction and relaxation in different smooth muscles like jejunum, trachea and aorta. So, we followed the protocols established by using different transducer systems. Isometric transducer for jejunum tissue, while isometric transducer for trachea and aorta.

Q. The authors have to provide details for methodologies for assays including anticholinesterase, lipoxygenase, antioxidant, antiemetic, analgesic and antipyretic activities

Ans. Methodology has been added in respective portion.

Q. The authors indicated in methods that quercetin was used as positive standard for anti-oxidant activity testing, but in results ascorbic acid is shown?

Ans. Quercetin was eliminated from text and now ascorbic acid has been incorporated.

Q. Authors stated that they cited methods with minor modifications, but they did not indicate what is modified?

Ans. Such statement has been eliminated.

Q. In testing for anticholinesterase, lipoxygenase, antioxidant activities (Table 1, 2), why there is not negative control?

Ans. “Ethanol was taken as negative control” Mentioned in the methodology now.
Q. For antipyretic activities assays, what was the name and dose pyrogen administered?

Ans. An i.p. injection (0.5 ml/kg) of pyrogen (brewer yeast)

Q. Did the authors compared all results with controls for their statistical significances?

Ans. Yes

Referee 2

As recommended by second reviewer, we modified our manuscript title and also double checked small mistakes.

Kindly acknowledge receipt of this manuscript. I am available if there are any further queries.

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Highest Regards

Prof. Dr. Vincenzo De Feo

Dipartimento di Scienze Farmaceutiche e Biomediche,

Università degli Studi di Salerno, Italy

E-mail: defeo@unisa.it