**Supplemental experimental procedures**

**Phosphorylation of the Tau protein**

The CDK2/CycA3 protein was prepared as primarily described in (1) and modified in (2). Phosphorylation of Tau by the CDK/CycA3 kinase is described in (2). The ERK kinase was prepared as described in (3). 100µM of 15N-labelled recombinant Tau[165-245] was phosphorylated by incubation with 1 µM of activated ERK at 37°C during 3h in 200µL of phosphorylation buffer 50mM Hepes pH 8.0, 50 mM NaCl, 12.5 mM MgCl₂, 2.5mM ATP, 2mM DTT, 1 mM EDTA, 2mM EGTA and protease inhibitor cocktail (ROCHE, Complete Inhibitors without EDTA).

Rat brain extract was prepared by homogenizing rat brains (about 2g in 5 ml) in homogenizing buffer (10mM Tris.Cl pH 7.4, 5mM EGTA, 2mM DTT, 1µM okadaic acid (Sigma) supplemented with protease inhibitor cocktail. Ultra-centrifugation was next performed at 100,000g for 1 hour. The phosphorylation reaction of 10 µM 15N-Tau protein (4) was performed at 37°C for 24h with 500 µl of rat brain extract in 10 mL of phosphorylation buffer (2 mM ATP, 40mM Hepes.KOH pH 7.3, 2mM MgCl₂, 5mM EGTA, 2mM DTT complemented with a protease inhibitor cocktail and 1 µM okadaic acid (Sigma).

Enzymatic incubations were terminated by heating at 75°C fro 15 min and followed by centrifugation. The phosphorylation mixture was buffer-exchanged using desalting centrifugal devices (0.5ml bed of G25 resin, cut-off of 7KDa, Thermo Scientific Zeba Desalting Columns) against NMR buffer.

**Co-localization measurement**

Neurons were imaged with a Zeiss LSM 710 confocal imaging system using a 40x or 63x oil-immersion objectives. Z-stacks were 0.36 µm per step with a 1 AU pinhole. The co-localization coefficients were measured for each stack using ZEN 2012 software following the method already published by Manders et al (5).

**Supplemental References**

