Additional Material and Methods

cDNA synthesis and RT-qPCR. Total RNA was extracted using TriReagent solution (#AM9738, Life Technologies) according to the manufacturer’s instructions. 1ug RNA was used to synthesize cDNA using M-MLV reverse transcriptase (#M1701, Promega), and real-time PCR analysis was performed with ABI PRISM® 7900HT Sequence Detection System (#4317596, Applied Biosystems®) using SYBR® Green master mix (#4472908, Life Technologies) and gene-specific forward or reverse primers. The sequences and concentrations used for each primer pair were as follows: AURKA (200 nM) SeqFw: GAAAGCCGGAGTGGAGCAT, SeqRv: TGCCGAAGGTGGGACTGTAT; AURKB (200 nM) SeqFw: TGTCACCCCACCTGCACCTTTG, SeqRv: CAGCTGTGGGCTGGACATT; KRAS (200 nM) SeqFw: CCCAGGTGCGGGAGAGA; SeqRv: CAGCTCAACTACCACAAGTTT; GAPDH (800 nM) SeqFw: GAGCCGCATCTTCTTTTG, SeqRv: CCATGGGTCTGAGCGATGT; GUSB (200 nM) SeqFw: CTCATTTGGAATTTTGCCGATT, SeqRv: CCGAGTGAAGATCCCCGTGTTTTT; ACTB (400 nM) SeqFw: GGCACCCAGCACATGAAG, SeqRv: CCGATCCACACGGGAGTACTTG. Relative quantitation was determined by the $\Delta \Delta C_t$ method using GUSB, GAPDH or ACTB as endogenous controls.

3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide- (MTT)-reduction based viability assay. 1 x $10^3$ cells were seeded in 96-well adherent plates. After drug treatment, cells were incubated with 1 mg/mL MTT (#M5655, Sigma-Aldrich) for 2 hours and formazan crystals were then resuspended in DMSO. The reduction of MTT
to formazan was measured colorimetrically at 570 nm in a EON™ plate reader (Biotek®).

All conditions were done in octuplicate.