A. Collect plants (50 mg in 500 µl RNA later)
   Store overnight at 4°C, then at -20°C

B. Transfer plants to 350 µl RA1 buffer (containing 1% β-mercaptoethanol)
   Add stainless steel cone ball
   Lyse plants using TissueLyser (25 s⁻¹ for 2x 2 mins)
   Centrifuge at 11,000x g for 5 mins

C. Transfer s/n to filter plate (approx 300 µl)
   Centrifuge at 3,200x g for 10 mins
   Move 150 µl s/n to deepwell plate
   Store other 150 µl s/n at -20°C

D. Add 150 µl RA4 buffer to lysed plant sample in deepwell plate
   Mix by pipetting 150 µl up and down x12
   Transfer 300 µl to binding plate

E. Pass solution through the binding plate by vacuum (-200 mbar, 2x 60 secs) or centrifuge (3,220xg, 2 mins)
   Add 460 µl RA3 buffer
   Pass solution through the binding plate by vacuum (-200 mbar, 2x 60 secs) or centrifuge (3,220xg, 2 mins)
   Add 29 µl DNasel solution and incubate for 10 mins
   Add 460 µl RA2 buffer
   Pass solution through the binding plate by vacuum (-200 mbar, 2x 60 secs) or centrifuge (3,220xg, 2 mins)
   Add 800 µl RA3 buffer
   Pass solution through the binding plate by vacuum (-200 mbar, 2x 60 secs) or centrifuge (3,220xg, 2 mins)
   Add 460 µl RA4 buffer
   Pass solution through the binding plate by vacuum (-200 mbar, 2x 60 secs) or centrifuge (3,220xg, 2 mins)

F. Dry membrane by centrifugation at 3,220xg for 10 mins
   Place binding plate over elution plate
   Add 50 µl RNase-free water to binding plate
   Incubate at RT for 3 mins
   Centrifuge plate sandwich at 3,220x g for 3 mins
   Purified RNA in elution plate