SI Fig. 2: Representative gel to confirm the presence of arbuscular mycorrhizal fungal DNA in all samples used for qPCR in both Plant and Fungal Diversity experiments. In the gel shown above, lanes 1-8 in both rows represent individual samples from the Fungal Diversity experiment. The presence of AM fungal DNA was confirmed using general PCR amplification with the primers AML1 and AML2 (Lee et al. 2008) using the Veriti thermal cycler PCR System (Applied Biosystems) in 8-well strip microtubes. Each 20µL reaction mixture PCR reaction contained 12µL of ddH2O, 2.50µL of buffer (10x), 0.30µL of BSA, 0.20µL of dNTPs, 0.50µL of AML1 forward primer (25µM), 0.50µL of AML2 reverse primer (25µM), 1.50µL of magnesium chloride, 0.50µL of Taq and 2.00µL of DNA. Cycling conditions were as follows: 95°C for 5 minutes; 30 cycles: 95°C for 45 seconds, 50°C for 1 minute, 72°C for 45 seconds; 72°C for 10 minutes and 4°C infinitely. Amplification for samples in the Fungal diversity experiment followed the same protocol as above with the following modifications to the PCR recipe: each PCR reaction contained 12.5µL of ddH2O, 2.50µL of buffer (10x), 0.30µL of BSA, 0.20µL of dNTPs, 0.25µL of AML1 forward primer (25µM), 0.25µL of AML2 reverse primer (25µM), 1.50µL of magnesium chloride, 0.50µL of Taq and 2.00µL of DNA. Cycling conditions remained the same.