

1 Applied Microbiology and Biotechnology

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3 **Copper Tolerance in *Frankia* sp. Strain Eu11c involves Surface-**
4 **Binding and Copper Transport.**

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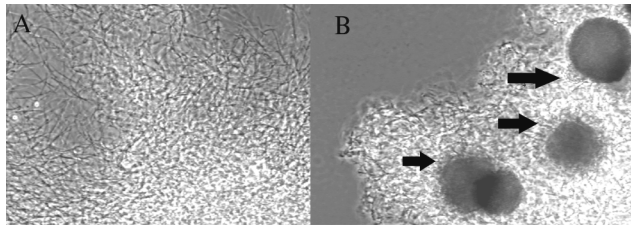
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19 Table S1 Primers used in this study

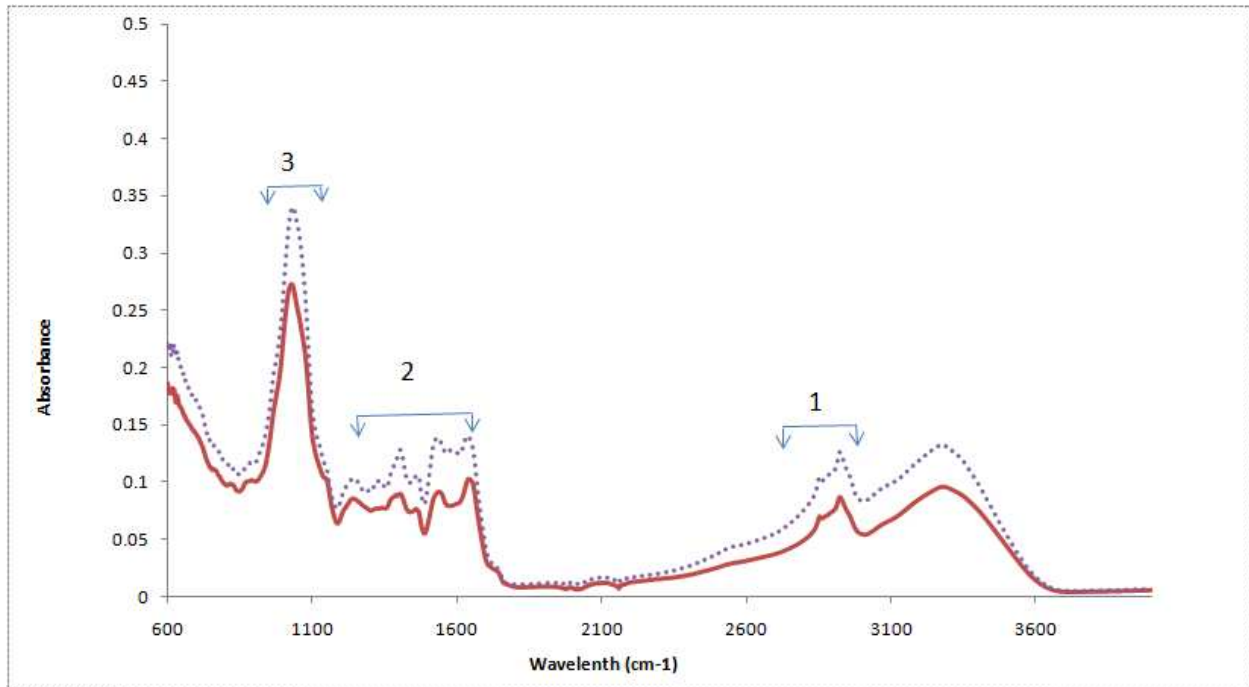
Locus ID (gene name)	Forward (5' → 3') primer Reverse (5' → 3') primer	Description
FraEu11c_6235	F-AACACCACCGGTGAGATCAAGA R-ATCACCTTGCAGGAGAACAGGT	rpoB
FraEu11c_4907	F-AGCTGGACAAGAACCGCAACAA R-AAGTTGACGATGGAGCTGACGA	rpsA
FraEu11c_0940	F-TCGGCAAGACCGTCATCATCCA R-ATCACCTTGCAGGAGAACAGG	atpD
FraEu11c_4734	F-TCTACCGCCTGTCCTACCGGAT R-ACGAACCGGACCCTACGTCA	copC1
FraEu11c_7109	F-TCTACAAGCTGTGGTTCGTGCT R-ATCACCCACCGGTTTCGACAGGAA	copCD
FraEu11c_1869	F-TCCCAGCTGTTACCACCATGTA R-CGAAGATGAGGATCTTCACGCCGA	copD
FraEu11c_6307	F-TTCTGGGCGTTCGCCTACAACG R-ATCACCTTGCAGGAGAACAGGT	copA (copper-translocating P-type ATPase)
FraEu11c_6308	F-GTATGACCTGCGGGCACTGTGT R-TGGTAGCCGGCCTCTTCGACGG	copZ (Heavy metal transport/ detoxification protein)
FraEu11c_7040	F-TCCTGATGATCACCAACCTGGACT R-AAGCTGACGTAGTACTGAGCGGAA	Periplasmic Binding protein/lacI
FraEu11c_4628	F-ACACCAAGAAGAAGTCCGTCACCA R-GATCGTGATGCTGTTGTCGGCATA	Hypothetical protein
FraEu11c_6772	F-TGACCAAGCAGATCAAGTTCACGC R-TTCACCTGGTAGATGTAGAACGACGC	ABC transporter ligand- binding protein
FraEu11c_1092	F-AAGGTCAACAAGGAGTTCTTCGACGC R-TCATCCGGTAGCGTTCTCGATCTT	Sulfate ABC transporter, periplasmic sulfate-binding protein

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23 Figure S1. Photomicrograph of *Frankia* strain DC12 grown in growth medium supplemented
 24 with (B) and without Cu^{+2} (A). Arrow points to globular structure on surface of hyphae.
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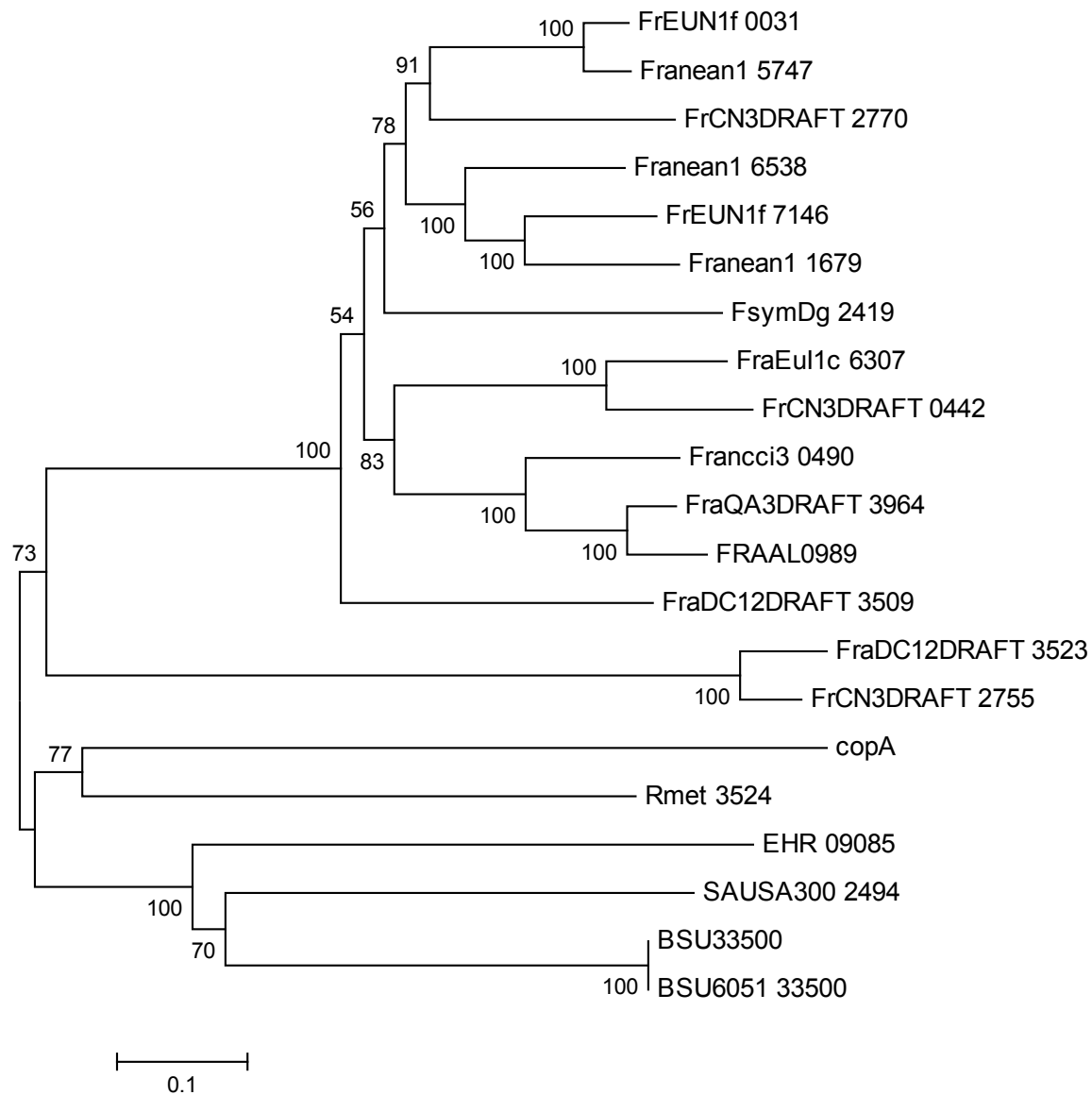


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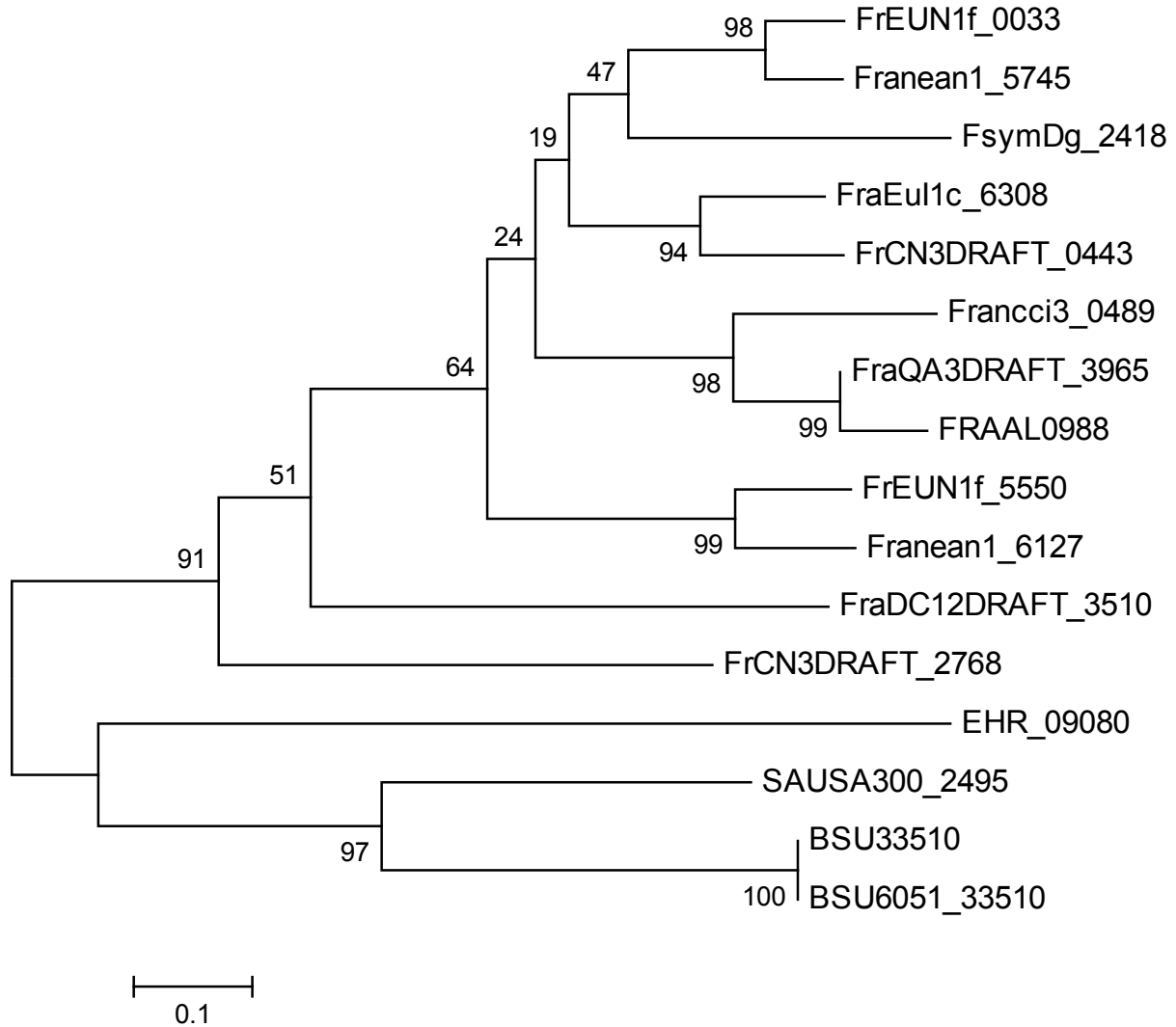
27 Figure S2. FTIR spectra of Cu^{+2} -stressed *Frankia* cells. Cultures were incubated for 5 days in
 28 low phosphate growth medium with (dashed line) or without (solid line) 1 mM CuSO_4 . After
 29 incubation, the cultures were collected and treated as described in the Methods. FTIR scans of
 30 lyophilized cells were taken and the averaged scans are presented (n=3). Numbers in figure
 31 represents areas of change and correspond to specific chemical signatures: (1) fatty acids, (2)
 32 fatty acids - proteins and phosphate-carrying compounds, and (3) cell wall carbohydrates –
 33 polysaccharides – PO_4 groups.
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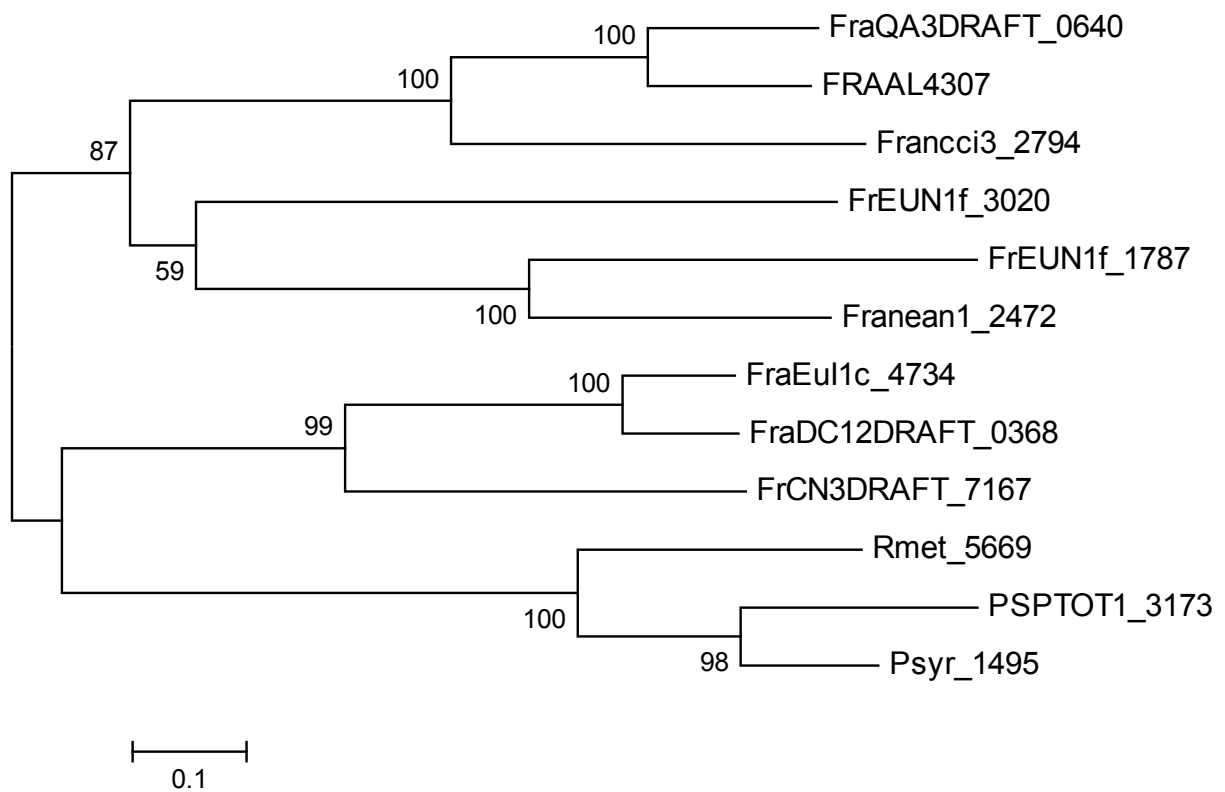


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 37 Figure S3. Phylogenetic tree of CopA protein sequences. The evolutionary history was inferred
 38 using the Neighbor-Joining method (Saitou and Nei 1987). The optimal tree with the sum of
 39 branch length = 5.35347782 is shown. The percentage of replicate trees in which the associated
 40 taxa clustered together in the bootstrap test (1000 replicates) are shown next to the branches
 41 (Felsenstein 1985). The tree is drawn to scale, with branch lengths in the same units as those of
 42 the evolutionary distances used to infer the phylogenetic tree. The evolutionary distances were
 43 computed using the JTT matrix-based method (Jones et al. 1992) and are in the units of the
 44 number of amino acid substitutions per site. The analysis involved 21 amino acid sequences. All
 45 ambiguous positions were removed for each sequence pair. There were a total of 1106 positions
 46 in the final dataset. Evolutionary analyses were conducted in MEGA5 (Tamura et al. 2011).
 47 Besides the *Frankia* CopA protein sequences, the following protein sequences were used:
 48 *Escherichia coli* K12 W3310 (copA), *Enterococcus hire* (EHR_09085), *Bacillus subtilis* strain
 49 168 (BSU33500), *Bacillus subtilis* 6051HGW (BSU6051_33500), *Staphylococcus aureus*
 50 USA300 SAUSA300_2494, and *Cupriavidus metallidurans* Rmet_3524).



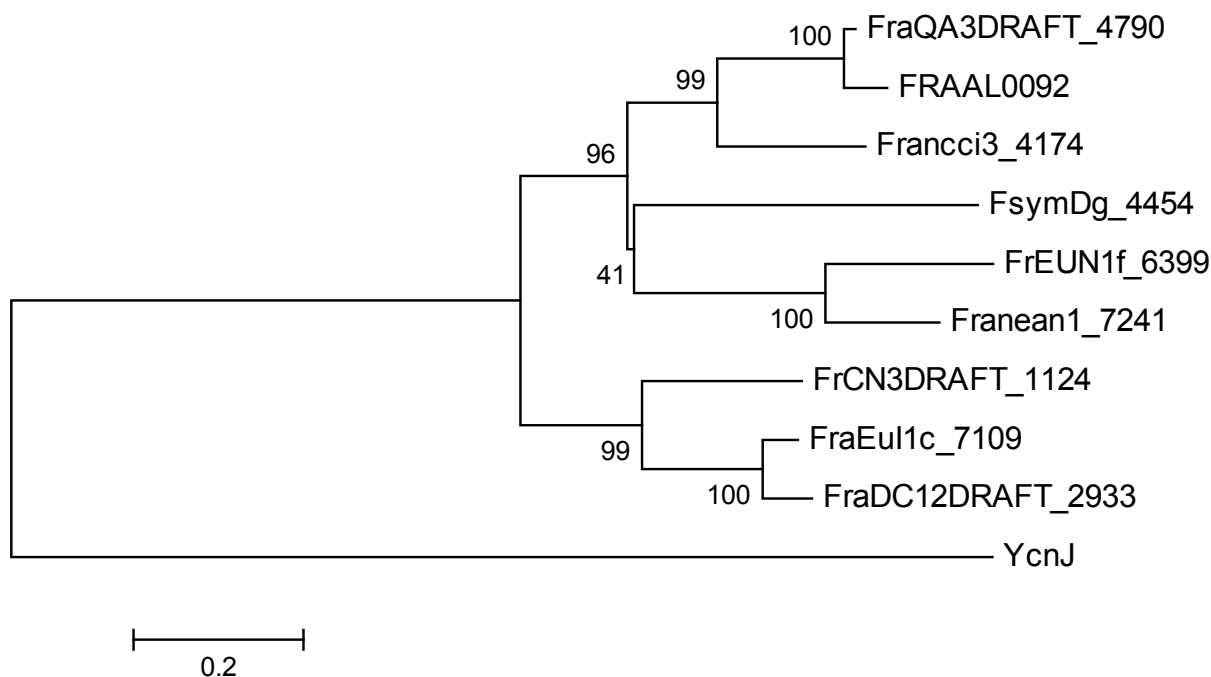
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53 Figure S4. Phylogenetic tree of CopZ protein sequences. The evolutionary history was inferred
 54 using the Neighbor-Joining method (Saitou and Nei 1987). The optimal tree with the sum of
 55 branch length = 4.84389874 is shown. The percentage of replicate trees in which the associated
 56 taxa clustered together in the bootstrap test (1000 replicates) are shown next to the branches
 57 (Felsenstein 1985). The tree is drawn to scale, with branch lengths in the same units as those of
 58 the evolutionary distances used to infer the phylogenetic tree. The evolutionary distances were
 59 computed using the JTT matrix-based method (Jones et al. 1992) and are in the units of the
 60 number of amino acid substitutions per site. The analysis involved 16 amino acid sequences. All
 61 ambiguous positions were removed for each sequence pair. There were a total of 82 positions in
 62 the final dataset. Evolutionary analyses were conducted in MEGA5 (Tamura et al. 2011).
 63 Besides the *Frankia* CopZ protein sequences, the following protein sequences were used: *E. hire*
 64 (EHR_09080), *B. subtilis* strain 168 (BSU33510), *B. subtilis* 6051HGW (BSU6051_33510), and
 65 *S. aureus* USA300 (SAUSA300_2495).
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68 Figure S5. Phylogenetic tree of CopC protein sequences. The evolutionary history was inferred
 69 using the Neighbor-Joining method (Saitou and Nei 1987). The optimal tree with the sum of
 70 branch length = 5.05269378 is shown. The percentage of replicate trees in which the associated
 71 taxa clustered together in the bootstrap test (1000 replicates) are shown next to the branches
 72 (Felsenstein 1985). The tree is drawn to scale, with branch lengths in the same units as those of
 73 the evolutionary distances used to infer the phylogenetic tree. The evolutionary distances were
 74 computed using the JTT matrix-based method (Jones et al. 1992) and are in the units of the
 75 number of amino acid substitutions per site. The analysis involved 12 amino acid sequences. All
 76 ambiguous positions were removed for each sequence pair. There were a total of 287 positions in
 77 the final dataset. Evolutionary analyses were conducted in MEGA5 (Tamura et al. 2011).
 78 Besides the *Frankia* CopC protein sequences, the following protein sequences were used: *C.*
 79 *metallidurans* Rmet_3524), *Pseudomonas syringae* *pv.* *tomato* (PSPTOT1_3173), *Pseudomonas*
 80 *syringae* *pv.* *syringae* B728a (Psyr_1495).

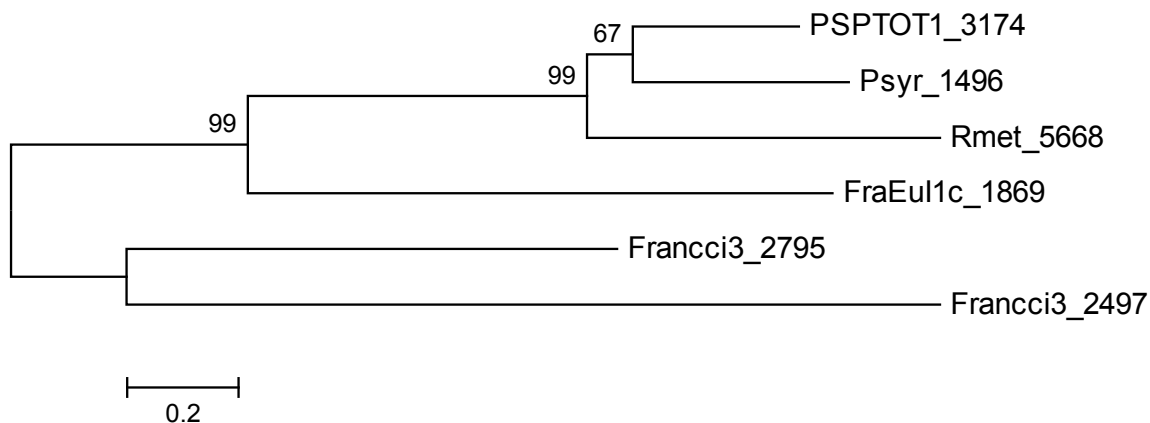


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82 Figure S6. Phylogenetic tree of CopCD protein sequences. The evolutionary history was
 83 inferred using the Neighbor-Joining method (Saitou and Nei 1987). The bootstrap consensus tree
 84 inferred from 1000 replicates is taken to represent the evolutionary history of the taxa analyzed
 85 (Felsenstein 1985). Branches corresponding to partitions reproduced in less than 50% bootstrap
 86 replicates are collapsed. The percentage of replicate trees in which the associated taxa clustered
 87 together in the bootstrap test (1000 replicates) are shown next to the branches. The evolutionary
 88 distances were computed using the JTT matrix-based method (Jones et al. 1992) and are in the
 89 units of the number of amino acid substitutions per site. The analysis involved 10 amino acid
 90 sequences. All ambiguous positions were removed for each sequence pair. There were a total of
 91 860 positions in the final dataset. Evolutionary analyses were conducted in MEGA5 (Tamura et
 92 al. 2011). Besides the *Frankia* CopCD protein sequences, the *B. subtilis* 6051HGW (YcnJ) was
 93 used.

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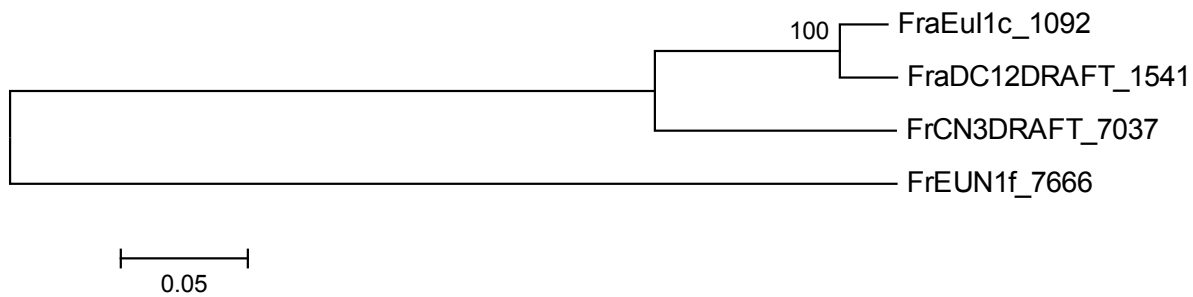
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97 Figure S7. Phylogenetic tree of CopD protein sequences. The evolutionary history was inferred
 98 using the Neighbor-Joining method (Saitou and Nei 1987). The optimal tree with the sum of
 99 branch length = 6.04157447 is shown. The percentage of replicate trees in which the associated
 100 taxa clustered together in the bootstrap test (1000 replicates) are shown next to the branches
 101 (Felsenstein 1985). The tree is drawn to scale, with branch lengths in the same units as those of
 102 the evolutionary distances used to infer the phylogenetic tree. The evolutionary distances were
 103 computed using the JTT matrix-based method (Jones et al. 1992) and are in the units of the
 104 number of amino acid substitutions per site. The analysis involved 6 amino acid sequences. All
 105 ambiguous positions were removed for each sequence pair. There were a total of 767 positions in
 106 the final dataset. Evolutionary analyses were conducted in MEGA5 (Tamura et al. 2011).
 107 Besides the *Frankia* CopD protein sequences, the following protein sequences were used: *C.*
 108 *metallidurans* Rmet_5668), *P. syringae pv. tomato* (PSPTOT1_3174), *P. syringae pv. syringae*
 109 B728a (Psyr_1496).

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114 Figure S8. Phylogenetic tree of the sulfate-binding periplasmic transport protein sequences. The
115 evolutionary history was inferred using the Neighbor-Joining method (Saitou and Nei 1987). The
116 optimal tree with the sum of branch length = 0.81330031 is shown. The percentage of replicate
117 trees in which the associated taxa clustered together in the bootstrap test (1000 replicates) are
118 shown next to the branches (Felsenstein 1985). The tree is drawn to scale, with branch lengths in
119 the same units as those of the evolutionary distances used to infer the phylogenetic tree. The
120 evolutionary distances were computed using the JTT matrix-based method (Jones et al. 1992)
121 and are in the units of the number of amino acid substitutions per site. The analysis involved 4
122 amino acid sequences. All ambiguous positions were removed for each sequence pair. There
123 were a total of 387 positions in the final dataset. Evolutionary analyses were conducted in
124 MEGA5 (Tamura et al. 2011).

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126 **References:**

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